RECORDS ADMINISTRATION

ACC#726759 DP-MS-78-71

SRL
ISOLATION OF NAEGLERIA FOWLERI FROM ARTIFICIALLY HEATED WATER CORD COPY

bу

C. B. Fliermans Savannah River Laboratory E. I. du Pont de Nemours & Co. Aiken, SC 29801

R. L. Tyndall University of Tennessee Knoxville, TN 37831

E. J. P. Willaert Medical Research Institute Veterans Administrative Hospital Gainesville, FL 32602

A paper proposed for publication in Nature as a letter.

This paper was prepared in connection with work under Contract No. AT(07-2)-1 with the U.S. Department of Energy. By acceptance of this paper, the publisher and/or recipient acknowledges the U.S. Government's right to retain a nonexclusive, royalty-free license in and to any copyright covering this paper, along with the right to reproduce and to authorize others to reproduce all or part of the copyrighted paper.

This document was prepared in conjunction with work accomplished under Contract No. DE-AC09-76SR00001 with the U.S. Department of Energy.

DISCLAIMER

This report was prepared as an account of work sponsored by an agency of the United States Government. Neither the United States Government nor any agency thereof, nor any of their employees, makes any warranty, express or implied, or assumes any legal liability or responsibility for the accuracy, completeness, or usefulness of any information, apparatus, product or process disclosed, or represents that its use would not infringe privately owned rights. Reference herein to any specific commercial product, process or service by trade name, trademark, manufacturer, or otherwise does not necessarily constitute or imply its endorsement, recommendation, or favoring by the United States Government or any agency thereof. The views and opinions of authors expressed herein do not necessarily state or reflect those of the United States Government or any agency thereof.

This report has been reproduced directly from the best available copy.

Available for sale to the public, in paper, from: U.S. Department of Commerce, National Technical Information Service, 5285 Port Royal Road, Springfield, VA 22161, phone: (800) 553-6847, fax: (703) 605-6900, email: orders@ntis.fedworld.gov online ordering: http://www.ntis.gov/ordering.htm

Available electronically at http://www.doe.gov/bridge

Available for a processing fee to U.S. Department of Energy and its contractors, in paper, from: U.S. Department of Energy, Office of Scientific and Technical Information, P.O. Box 62, Oak Ridge, TN 37831-0062, phone: (865) 576-8401, fax: (865) 576-5728, email: reports@adonis.osti.gov

ISOLATION OF NAEGLERIA FOWLERI FROM ARTIFICIALLY HEATED WATER*

C. B. Fliermans Savannah River Laboratory E. I. du Pont de Nemours & Co. Aiken, SC 29801

R. L. Tyndall University of Tennessee Knoxville, TN 37831

E. J. P. Willaert Medical Research Institute Veterans Administrative Hospital Gainesville, FL 32602

Power generating systems in the United States will probably reach 1.3×10^6 megawatts by 1990 requiring an additional 7.5×10^6 gpm of cooling water. The discharge of any waste heat from industralization into southeastern aquatic habitats results in the heating of already warm water, $25-35^{\circ}C.^{2}$, To determine if such thermal discharges into warm waters enhance the isolation frequency of Naegleria fowleri, we compared a variety of aquatic habitats altered by man-made thermal discharges at the Savannah River Plant (SRP), near Aiken, South Carolina. Additionally, control samples were collected from ambient temperature lakes, ponds, the Savannah River, and the unheated artificial streams at the Flowing Streams Laboratory (FSL). Of the 79

^{*} The information contained in this article was developed during the course of work under Contract No. AT(07-2)-1 with the U.S. Department of Energy.

samples from artificially heated habitats, 55 demonstrated outgrowth at 45°C and 34 had flagellated amoebae characteristic of N. fowleri. 5 Only one high temperature flagellated amoeba was isolated from the 51 samples collected from ambient temperature habitats. Our data indicate that N. fowleri is more frequently isolated from thermally altered habitats, and that the isolation frequency increase where habitat temperatures are between 40° and 60°C.

Selected free-living species of Acanthamoeba and Naegleria (N. fowleri) are causative agents of primary amoebic meningoencephalitis (PAM).^{6,7} The true incidence of the disease is probably unknown because many cases are either unrecognized or incorrectly diagnosed; however over 150 cases of fatal PAM have been attributed to N. fowleri and Acanthamoeba. Subsequent epidemiological studies indicated that many cases of PAM occurred in individuals who had been in close contact with heated water.^{8,9}

Griffin 10 demonstrated that N. fowleri and A. culbertsoni
grew in vitro at temperatures above 37°C while their nonpathogenic
counterparts did not, and suggested that thermal additions to
aquatic systems may select for pathogenic species of the ubiquitous
free-living amoebae. De Jonckheere, et al. 5 noted that in Belgium,
N. fowleri has been isolated only from artificially heated water.
Recent studies in the U.S. also demonstrated that N. fowleri
isolated more frequently from southeastern aquatic systems during the
summer months. 2,11,12

In this study, water, sediment, and algal samples were collected from both thermal and ambient temperature systems throughout the 777 square kilometer (SRP) site. The majority of samples were collected from either the Par Pond system³ or from artificially heated streams at the FSL. 4 The Par Pond system consists of a 1092 hectare cooling reservoir which is a source of 90% of the water used to cool a production reactor. The resulting thermal effluent (ca 768,000 1/min) is discharged to the environment through a series of canals and cooling ponds, 11 kilometers long, before being discharged again into Par Pond. Within the discharge canal system, thermal gradients are established with corresponding algal and bacterial communities. The second major habitat, FSL, is a greenhouse facility containing four artificially heated streams. Water from an unheated blackwater stream, Upper Three Runs Creek, is heated to a temperature of 25°C above ambient and then mixed with increasing amounts of ambient temperature water to achieve streams of 12.5°, 7.5°, 5.0°, and 2.5°C above ambient. Additionally, two control streams are maintained with ambient temperature water.

All 130 samples were collected aseptically and processed immediately in the field or within 24 hours after returning to the laboratory. Temperature, pH, and dissolved oxygen were measured at the time of collection. Water samples were vacuum filtered through sterile cellulose acetate membrane filters (1.2 nm) at 25.4 cm of mercury. The filters were removed, aseptically cut

in half, and inverted onto nonnutrient agar plates seeded with heat-killed and unheated *Escherichia coli*. Algal communities, microbial mats, and sediments were placed directly on *E. coli* seeded plates. Plates were sealed with parafilm, incubated at 45°C, and observed daily for outgrowth. When sufficient outgrowth had occurred, amoebae were harvested by washing with sterile water and transferred to newly prepared *E. coli* seeded plates. Stock cultures were maintained on *E. coli* seeded plates or axenically in a casitone-based medium. 13

Amoebic isolates were identified as N. fowleri on the basis of their ability to grow at 45°C, their typical trophozoite and cyst morphology, and their ability to flagellate. Additionally, serological analyses were made using indirect immunofluorescence with antisera prepared against the pathogen N. fowleri5 and the nonpathogenic Naegleria, i.e., N. jadini and N. gruberi. The isolates reacted maximally with antisera against pathogenic N. fowleri (Morgan), although minimal reaction was noted with antisera against N. jadini and N. gruberi. Pathogenicity of the isolates was tested by intranasal inoculation of $8-16 \times 10^4$ trophozoites into ether-anesthetized 4-6 week 1CR or BALB/c mice. Some isolates produced fatal meningoencephalitis on intranasal inoculation and were readily reisolated from infected brain tissue, while other isolates produced no PAM. Serological analyses of selected nonpathogenic isolates confirmed them as N. fowleri, and are considered as nonpathogenic variants often associated with artificially heated water. 5,14

The data for studies conducted from January to May of 1978 are summarized in Table 1, and indicate that 43% of the samples taken from thermally altered habitats were positive for N. fowleri while ambient temperature samples were positive only 1.9% of the time. The data further indicate that N. fowleri are more frequently isolated from habitats with elevated temperatures than cooler habitats (Figure 1). It is interesting to note that the heated streams at FSL had been operating only about 18 months when the first isolation of N. fowleri was made, while control streams have continued negative by our sampling techniques. This suggests that N. fowleri can colonize a thermally altered habitat rather quickly. Additionally, thermal systems at the site have existed since the early 1950's; thus, the presence of N. fowleri in these systems is probably not recent.

Our studies suggest that thermally altered aquatic systems may provide habitats that are conducive to the growth and proliferation of *N. fowleri*. The ecology of pathogenic micro-organisms associated with the receiving waters merits further study.

LITERATURE CITED

- United States Environmental Protection Agency. EPA 440/1-74,
 029-a (1974).
- 2. Wellings, F. M., Amuso, P. T., Chang, S. L., and Lewis, A. L. Appl. Environ. Microbiol. 34, pg 701-705 (1977).
- 3. Fliermans, C. B., Gorden, R. W., Hazen, T. C., and Esch, G. W. Appl. Environ. Microbiol. 33, pg 114-122 (1977).
- 4. Harvey, R. S. Assoc. Southeast Biological Bull. 20, pg 3-7 (1973).
- De Jonckheere, J., Van Dijck, P., and Van de Voorde, H.
 J. Hyg. (Cambridge) 75, pg 7-13 (1975).
- Culbertson, C. D., Smith, J. C., Cohen, H., and Minner, J.
 Amer. J. Path. 35, pg 185-187 (1959).
- 7. Fowler, M., and Carter, R. F. Brit. Med. J. 2, pg 740-742 (1965).
- 8. Duma. R. J. Crit. Rev. Clin. Lab. Sci. 3, pg 163-201 (1972).
- 9. Willaert, E. Ann. Soc. Belge Med. Trop. 54, pg 205-216 (1974).
- 10. Griffin, J. L. Science 178, pg 869-870 (1972).
- 11. Stevens, A. R., Tyndall, R. L., Coutant, C. C., and Willaert, E. Appl. Environ. Microbiol. 34, pg 701-705 (1977).
- 12. Tyndall, R. L., Willaert, E., Stevens, A. R., and Coutant, C. C. Isolation of Pathogenic Naegleria from Artificially Heated Waters. Proceedings of Microbiology

- of Power Plant Effluents, University of Iowa, pg 117-124 (1977).
- 13. Willaert, E. Ann. Soc. Belge Med. Trop. 51, pg 701-708 (1971).
- 14. De Jonckheere, J., and Van de Voorde, H. J. Protozool. 24, pg 304-310 (1977).

TABLE 1
Summary Data for the Presence of Thermophilic Flagellated Amoeba at the Savannah River Plant

Habitat	Temperature, °C	Number of Sampl es	Samples With Outgrowth of Amoeba at 45°C	Samples With Flagellated Amoeba	Number of Flagellated Amoeba/ Number of Samples
Artificially heated	9.8 - 71.5	79	55	34	43.0
Ambient temperature	5.2 - 28.5	51	7	1	1.9

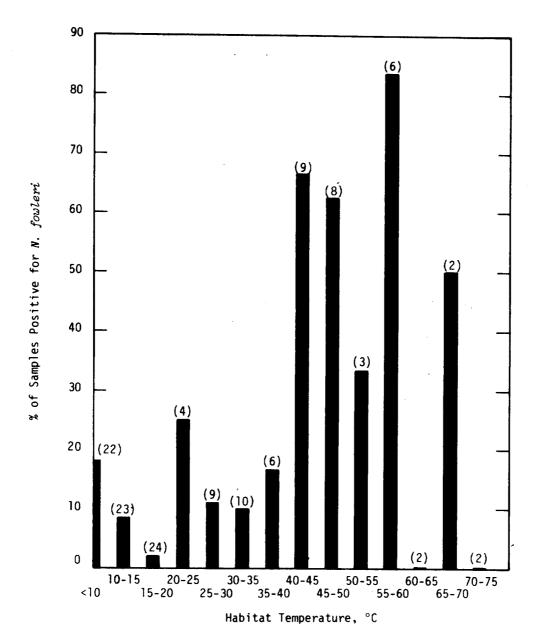


FIGURE 1. Percentage of Samples Positive for N. fowleri with Respect to Habitat Temperature (sample size in parentheses)